

INTRODUCTION

Characterization of *Enterococcus* Species in Coastal Waters Exceeding Water Quality Standards

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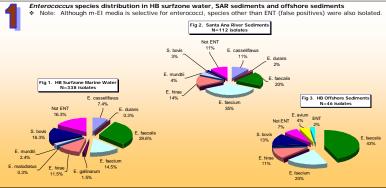




Fig 10. Genotypic Characterization of E. faecalis Isolates in Correlation to Geographic Location, Source and Biotype River Sediment Marin H:O 61 (42) 5 (3) 4(3) 86 (54) 1 (1) 6 (2) 143311 & 7143711 1 (1) 3 (2) 142711 & 7143711 5 (4) 5 (5) 7142711 5 (5) 2(2) 2(2) 7142711 2 (2) 2 (1) 7142711 2(1) 7143711 2(1) 1 (1) 1 (1) 2(2) 1 (1) 7143711 2 (2) 2 (1) 2(1) 3 (2) 7143711 3 (2) 2 (2) 1 (1) 1 (1) 1 (1) 1 (1) 1 (1) 7143711 3(3) 7143711 2(2) 2(1) 2 (1) 2(1) 7143711 2(1) 7143711 2(1) 7142711 4(3) 1 (1) 1 (1) 2(1) 3(1) 7143711 5(2) 3 (2) 7143711 3 (2) 2 (1)

Populations of E. faecalis that appear to be clonal were found in water, sediments and gull stools from Huntington Beach as

well as Baby Beach, located 20 miles south. The majority of clones had the same PFGE type (I) and included isolates that

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2(2)

LEGEND

HB - Huntington Beach NB – Newport Beach

SAR - Santa Ana River BB - Baby Beach

SD - Storm Drain

STUDY OBJECTIVES -Identify ENT species distribution in marine water and

Huntington Beach

1-Identify ENT species distribution...... fecal associated species and to identify predominant species for Pulsed-Field Gel Electrophoresis (PFGE) typing. n-Determine levels of FIB in sediments from locations

A section of Huntington Beach (Fig. 1), California, has frequently been posted or

closed for swimming since 1999, primarily due to high levels of enterococci fecal

indicator bacteria at the surf zone. Numerous multi-disciplinary investigations have

not identified specific sources responsible for the beach water quality failures. A bacteriological study was conducted to characterize enterococci species distribution

at 4 surf zone and 2 river mouth sites for samples that passed or failed water quality

standards. Samples were collected 4-6 days/week for a 6-week period that included 3 spring tide cycles. Enterococci isolates (N=338) were obtained using mEI media (EPA

Method 1600) and up to 5 isolates/sample were speciated using API 20 STREF

(Biomérieux, France) with additional biochemical testing. Enterococci single sample standards (> 104 CFU/100 ml) were exceeded on 21 of 31 sampling days with the

highest levels found at the surf zone sites during spring tides. During spring tide cycles, 33% of samples (N=58) exceeded standards as compared to 16% (N=55)

during neap tides. The most common species found in coastal waters were E. faecalis (30%), S. bovis (16%), E. faecium (15%), E. hirae (12%), E. casseliflavus (7%) and

other species identified as non-enterococcus (15%). Enterococcus faecalis and

Streptococcus bovis were the predominant species at the surf zone sites when levels were both well above and below standards. E hirae and E. casseliflavus were more

commonly detected at the river sites. There were no significant changes in species

distribution overall, in samples either passing or failing standards at all of the sites.

These results suggest that these species are generally present in coastal waters at these surf zone and river mouth sites and that increased levels may be related to tidal and

Enterococcus (ENT) is one group of fecal indicator bacteria (FIB) used

by government agencies to determine whether beaches are safe for

swimming. Contributing sources of ENT include humans, animals, birds

sewage, and natural and urban runoff. Some species of ENT, such as E.

faecalis and F. faecium are commonly associated with fecal sources

However, E. casseliflavus, E. mundtii and E. gallinarum are more

commonly found in plants and soil. ENT may also accumulate, persist

and possibly replicate in marine sediments that, when resuspended, can be a source of ENT to overlying water. Thus, high levels of ENT that are

environmental in origin may give a false indication of fecal

At Huntington Beach, high levels of ENT appear to correlate with spring tide conditions, particularly in summer. Spring tides occur over a period of a few consecutive days before and after a full moon (Fig. 7). During

this time, the gravitational forces from the moon and sun are at a maximum, resulting in the highest exchange of ocean water flowing in and out of coastal outlets. Contaminants in the Santa Ana River (SAR), including bacterial-laden sediments, are flushed out to the ocean during ebb tides, possibly causing beach water quality failures.

The aim of this study was to investigate the sources of ENT to

suspected of impacting beach.

-Investigate relationship between beach failures and tidal

-Compare ENT species distribution in beach water meeting and failing single sample ENT standards (104 CFU/100 ml). F-Compare relatedness of ENT strains using Proc. to determine spatial distribution and growth, as indicated by clonal strains

MATERIALS & METHODS

Sample collection

Water samples were collected at the HB surfzone and SAR mouth 4-6 days/week for a 6-week period that included 3 spring tide cycles. Ocean bottom sediments were collected at depths from 10 to 71 m using a clamshell sampler. Surface sediments (top 3") in the SAR were collected into a sterile bottle. Sediments were collected from areas suspected of contributing bacteria to the beach (Fig. 5)

- A. Santa Ana River (SAR), above Pacific Coast Highway (PCH) B. SAR below PCH
- C. Huntington Beach Coast, near the AES power plant
- D. Huntington Beach Coast. 0.2 mi from shore at 10 m depth
- Orange County Sanitation District (OCSD) sewage outfall
- F. Newport Beach coast (control site)

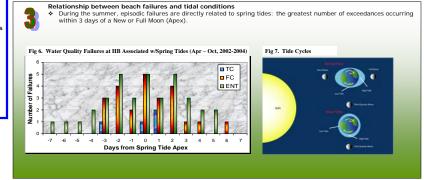
Enterococci levels in sediments and water Enterococci were extracted from sediment using sonication (30s at 30% output using a Branson Sonifier® 450) and enumerated using

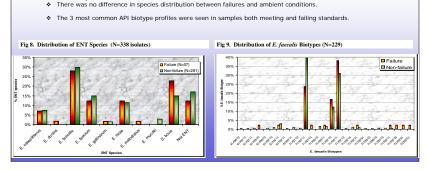
Membrane Filtration (FPA Method 1600). Enterococci speciation and distribution

Up to 5 isolates per sample were isolated from m-El media. Isolates were identified to *Enterococcus* species (biotype) level or as "non-enterococci" using API 20 STREP® and additional biochemical testing. The species distribution was determined by counting each species only once per sample.

Pulsed-field Gel Electrophoresis

E. faecalis strains were cultured in brain heart infusion broth for 18-24 h at 37°C and pelleted by centrifugation at 12,000g for 4 min. Bacterial cells were embedded in agarose plugs and lysed for 48 h at 37°C with EC lysis buffer and Mutanolysin stock. were washed and digested with Sma I endonuclease for 4 h at 25°C Electrophoresis was performed with a contour-clamped homogenou electric field device (CHEF-DRIII or CHEF MAPPER, Bio-Rad, CA) at 200V. 18h. Gels were stained with ethidium bromide (Bio-Rad. CA) and analyzed with BioNumerics program (Applied Maths, Kortrijk Belgium). A dendrogram was derived from the unweighted pair





Comparison of ENT species distribution in beach water meeting and failing single sample standards (104

Fig 11 PEGE Dendogram of E. faecalis Isolates

2 (2)

No. Clonal

were collected within a 2 month period.



DISCUSSION

Epidemiological studies have shown that swimming-related gastroenteritis is associated with ENT in fecal-contaminated coasts. All Huntington Beach, tidal events appear to correlate with increased levels of ENT, indicating bacterial input from a source that is tidally influenced. There was no significant change in species distribution when levels surpassed standards, suggesting persistent source(s) of ENT. We hypothesize that the SAR is one of these sources. Previous dye studies described how pollutants in the SAR could be transported to Huntington Beach. The high levels of ENT found in the SAR sediments indicate they may be replicating in this environment. The origin of ENT in sediments is still undetermined. The predominant species found in sediments are also common inhabitants of the intestinal microflora of humans and animals. The PFGE results suggest that there may be a clonal population of *E. faecalis* that is widely distributed in this coastal environment. It is possible that this "clonal" population is comprised of strains that are so closely related that additional restriction enzymes are needed for further discrimination. Future studies include using PFGE in conjunction with an additional typing technique for clonality assessment.

We thank the Orange County Sanitation District for providing mEI culture plates from water samples collected at the HB surfzone and Santa Ana River; Christopher Francis Justin Ignacio, Bonnie Steward, Rina Tijotahadi and Margaret Clarke for their technical assistanc



